

## PICKING BACTERIAL COLONIES AND LIQUID CULTURE

Est. Total Time: 1 Hour.

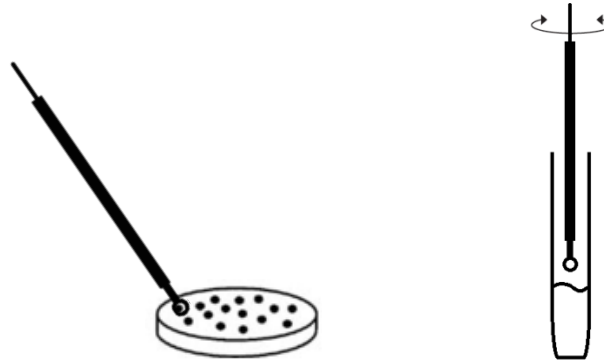
**Summary:** Use this technique to go from bacterial plates to liquid culture, or simply to grow more bacteria.

### **Materials**

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| <ul style="list-style-type: none"> <li>- 1L of autoclaved LB Broth, Terrific Broth, or selective liquid nutrient media</li> <li>- Bacterial Plates with single colonies of interest</li> <li>- 70% Ethanol or Isopropanol</li> <li>- Inoculation Loops and/or Pipet Tips</li> <li>- Pipet bulb or Autopipettor</li> <li>- 10 mL Serological Pipets</li> <li>- Snap cap Culture Tubes</li> <li>- A marker, preferably super-permanent.</li> </ul> | <ul style="list-style-type: none"> <li>- MaxQ Shaker Incubator w/ blue test tube rack</li> </ul> <p><b><i>OPTIONAL:</i></b></p> <ul style="list-style-type: none"> <li>- Ampicillin, (Cell Center)</li> <li>- Kanamycin,</li> <li>- Tetracycline,</li> <li>- X-Gal</li> <li>- S-Gal</li> </ul> |
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### **Procedure:**

1. Prepare your work area; wipe down the area down with 70% Ethanol or Isopropanol.
2. Obtain your bacterial plate(s) with the colony(ies) of interest on them.
3. Using the pipet bulb or autopipettor and the serological pipets, dispense 5 mL of liquid culture medium into the snap cap culture tubes. One tube per colony.
4. Label the tubes.
5. Using an inoculation loop or pipette and tip, gently pick a single colony from the plate and inoculate the corresponding culture tube, as in the diagrams below (submerge the loop!):



6. Cap the tubes to the 'first stop'.
7. Rack the culture tubes at an angle in blue test tube racks, and incubate at 37°C with shaking at 250rpm in the MaxQ shaker incubator. Check on your cultures the following day.

### **Cleanup:**

1. Replace all reagents in their proper locations.
2. Dispose of all waste.
3. Wipe down all surfaces used with 70% Ethanol or Isopropanol.
4. Consult Lab Staff if unsure about any of the above.